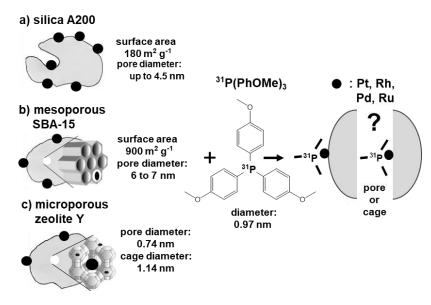
Method for the determining of the spatial distribution of noble metals on porous solids by complex formation with tris(methoxyphenyl)phosphine and ³¹P MAS NMR

Spectroscopic background: Due to the high reactivity of tris(methoxyphenyl)phosphine (P(PhOMe)₃) in the complex formation with noble metals (NM: Pt, Rh, Pd, Ru) and the molecular diameter of 0.97 nm, this molecule is suitable for determining the presence of noble metals at the outer particle surface of microporous zeolites, such as zeolite Y (structure type FAU), since the micropores and cages, such as the 12-ring pores of this zeolite with a diameter of 0.74 nm, are not accessible for this molecule. On amorphous silica (A200 or A300) and mesoporous SBA-15, on the other hand, all well-dispersed noble metal atoms are available for complex formation with P(PhOMe)₃ due to the open surfaces and large pores of these materials (see Fig. 1) [1].





Because of the natural abundance of 100% and a nuclear spin of $I = \frac{1}{2}$ of the ³¹P isotope, quantitative ³¹P MAS NMR spectroscopy is a suitable tool for the investigation of P(PhOMe)₃ complexes formed at noble metals on porous supports with high sensitivity. While bulk P(PhOMe)₃ causes a narrow ³¹P MAS NMR signal at $\delta_{31P} = -14$ ppm (Table 1 in [1]), physisorption of this molecule on open silica surfaces and in mesopores leads to a low-field resonance shift to $\delta_{31P} = -10$ ppm (**Fig. 2a** shown below and Fig. 1, right-hand side, in Ref. [1]). Same types of ³¹P MAS NMR signals are observed for adsorption of P(PhOMe)₃ on zeolite H-Y (**Fig. 2b** shown https://michael-hunger.de

below and Fig. 1, right-hand side, in Ref. [1]). The latter finding indicates that $P(PhOMe)_3$, in contrast to PPh_3 (see Fig. 2e in the Section "method 21"), is not able to enter the supercages of zeolite Y and to interact with the Broensted acidic bridging OH groups (Si(OH)AI) located in these cages.

³¹P HPDEC MAS NMR

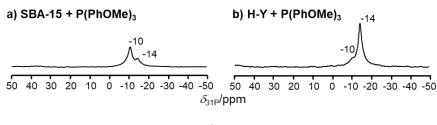


Fig. 2

For studying the complex formation of P(PhOMe)₃ with noble metals, H₂-reduced Pt-, Rh-, Pd-, and Ru-loaded meso- and microporous support materials were prepared as described in Refs. [1-5]. In their H₂-reduced state, these catalysts demonstrated a high activity in the hydrogenation of propene and the dehydrogenation of propane [4, 5]. The complex formation of P(PhOMe)₃ with noble metals, such as platinum and rhodium, on the silica A200 with a well accessible surface on mesoporous SBA-15 leads to broad ³¹P MAS NMR signals at δ_{31P} ca. 35 ppm, which is a similar chemical shift range as observed for complexes formed by adsorption of PPh₃ (compare leftand right-hand side of **Fig. 3** shown below and Fig. 2 in Ref. [1]).



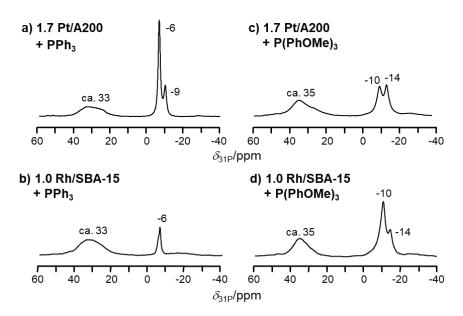
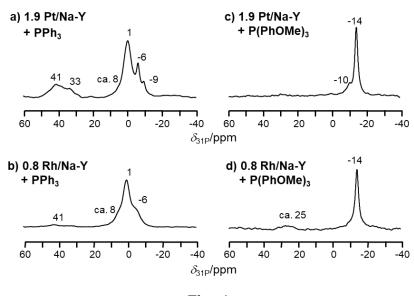


Fig. 3

In contrast, upon adsorption of P(PhOMe)₃ on platinum- and rhodium-loaded zeolites Na-Y, no ³¹P MAS NMR signals of complexes occur at δ_{31P} ca. 35 ppm, while complex formation with PPh₃ is indicated by signals at δ_{31P} ca. 33 and 41 ppm is indicated (compare left- and right-hand side of **Fig. 4** shown below and Fig. 3 in Ref. [1]). This observation additionally evidences that P(PhOMe)₃ is too large for forming complexes with noble metal atoms located in the pores and cages of zeolite Y. Since the pore and cage system of zeolite Y is characterized by 12-ring windows with a diameter of 0.74 nm, which is one of the largest of common microporous zeolites, also the complex formation of P(PhOMe)₃ with noble metal inside the pores and cages of other zeolites can be excluded.



³¹P HPDEC MAS NMR

Fig. 4

A summary of the above-mentioned ³¹P chemical shift values of bulk and physisorbed P(PhOMe)₃ as well as complexes of P(PhOMe)₃ with noble metals is given in Table 1 shown below. Considering these values, the **location of noble metals on the outer surface of zeolites can be determined by** formation of complexes with P(PhOMe)₃ leading to **signals at** δ_{31P} **ca. 35 ppm**. Contact of H₂-reduced noble metal-containing catalysts under study with oxygen (e.g. air) must be strictly avoided. Otherwise, oxidation of P(PhOMe)₃ can occur, leading to ³¹P MAS NMR signals with chemical shifts near the range of signals due to complex formation with noble metals (line 4 of Table 1).

$\delta_{ m 31P}$ / ppm	Assignment of P(PhOMe) ₃ species		
-14	bulk P(PhOMe) ₃		
-10	physisorbed P(PhOMe) $_3$ on open surfaces and in mesopores		
ca. 25	decomposed oxidation products of P(PhOMe) ₃		
ca. 35	[P(PhOMe] _n NM complexes on open surfaces and in mesopores		

Table 1

For the quantitative evaluation of the ³¹P MAS NMR signal intensities of noble metal complexes with phosphines, the procedure described by Equ. (3) of Section "method 12" is utilized. As an intensity standard, a well-known aluminosphosphate-type zeolite, such as VPI-5, is suitable. In this case, the repetition times t_{rep} of the ³¹P MAS NMR measurements can be optimized by solely evaluating the ³¹P MAS NMR signals of phosphines complexed with noble metals, recorded with $t_{rep} = 20$ s, while the VPI-5 standard is recorded with $t_{rep} = 240$ s. Phosphines physisorbed at the support surfaces without noble metals in their vicinity have much higher T₁ relaxation times compared with phosphines involved in complex formation with noble metals.

If $P(PhOMe)_3$ is applied together with PPh_3 (see Section "method 21") as a molecular probe for the study of the spatial distribution of noble metals on zeolite Y, the noble metal locations summarized in **Table 2** (Table 4 in Ref. [1]) and visualized in **Fig. 5** can be distinguished by their complex formation with phosphines. Ref. [6] gives a survey on the above-mentioned topics.

Positions in Fig. 5	Noble metal location	Complexation with phosphines	³¹ P MAS NMR signal (ppm)
1	on outer particle surface	PPh_3 and	ca. 33 and 35
	of DeA-Y and Na-Y	P(PhOMe) ₃	
2	in secondary	PPh_3	ca. 33
	mesopores of DeA-Y		
3	in supercages (SII	PPh_3	41
	position) of Na-Y		
4	in sodalite cages (SII'	no	no
	position)		
5	in hexagonal prisms (SI	no	no
	position)		

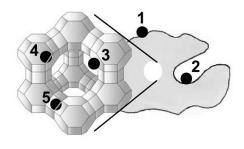


Fig. 5

Catalyst preparation: The noble metal-containing catalysts, prepared as described in Refs. [1-5], were calcined in synthetic air (970 mL/min) by heating with a rate of 2 K/min up to 573 K and further calcined at this temperature for 3 h, excluding the Rucontaining support materials. The materials were reduced in flowing hydrogen (100 mL/min) at 623 K (Pt-, Rh-, Pd-containing samples) or at 673 K (Ru-containing samples) for 2 h, transferred into glass tubes inside a glove box under nitrogen atmosphere, evacuated ($p < 10^{-2}$ Pa) at a vacuum line (see Section "vacuum line 1" via link "*In Situ* Solid-State NMR Techniques") at 298 K for 12 h, and sealed in glass tubes until their use.

For studies as those demonstrated in Figs. 2 to 4, loading of the H₂-reduced catalysts with tris(methoxyphenyl)phosphine (P(PhOMe)₃) of Sigma-Aldrich is performed by mixing of ca. 50 mg catalyst with ca. 10 mg P(PhOMe)₃ inside a rotor using e.g. a mini glove box purged with dry nitrogen gas (see Section "mini glove box" via link "*In Situ* Solid-State NMR Techniques"). Subsequently, the rotor is sealed with an O-ring-containing TORLON cap and heated at 433 K for 2 h for reaching a proper distribution of the P(PhOMe)₃ on the catalyst.

³¹*P* MAS NMR studies: ³¹P MAS NMR measurements were carried out using a Bruker Avance III 400WB spectrometer at a resonance frequency of 161.9 MHz using a 4 mm MAS NMR probe with sample spinning rates of 10 kHz. The spectra of the P(PhOMe)₃-loaded noble metal containing catalysts are recorded upon single pulse $\pi/2$ excitation with the repetition time of 20 s and high-power proton decoupling (HPDEC). The intensity standard VPI-5 is measured with a repetition time of 240 s. Chemical shifts are referenced to 0.85 M H₃P0₄ ($\delta_{31P} = 0$ ppm).

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