Characterization of dehydrated sodium-containing catalysts by ²³Na solid-state NMR

Spectroscopic background: ²³Na nuclei have a spin of I = 3/2 and a quadrupole moment of $Q = 10.4 \times 10^{-30} \text{ m}^2$. Therefore, ²³Na NMR signals of sodium atoms in dehydrated solids are affected by quadrupolar interactions. The ²³Na isotope has a natural abundance of 100 % and a sensitivity of 9.3 x 10^{-2} in comparison with ¹H nuclei (1.0), making this isotope a very suitable candidate for NMR studies of solids. For basic principles of solid-state NMR, see lectures "Solid-State NMR Spectroscopy" for Bachelor students or PhD seminars, accessible via the link "Lectures for Students".

If a catalyst framework contains atoms, which cause negative framework charges, these charges are compensated by extra-framework cations, such as Li⁺, Na⁺, Cs⁺, Ca²⁺, Mg²⁺ etc. Extra-framework cations in zeolites are coordinated with framework oxygen atoms and located on well-defined cation sites of the zeolite structure (see Fig. 1 for the faujasite structure of zeolites X and Y [1]). At ambient temperature, exchange of extra-framework cations between different crystallographically non-equivalent sites can be neglected on the time scale of NMR spectroscopy. In this case, the solid-state NMR spectra, e.g. of sodium cations, are mainly determined by the strength of their guadrupolar interactions, making this method a powerful tool for structure studies [1-10]. The quadrupole coupling constants, C_Q, depend on the electric field gradients, which in a first approximation are caused by a superposition of the electrostatic fields of the negatively charged framework oxygen atoms. Simply, the C_Q value is large for sodium cations, which are unsymmetrically coordinated to oxygen atoms, e.g.



Fig. 1

outside of the center of a 6-membered oxygen ring (positions SII in zeolite Y, see **Fig. 1**) and **small for a symmetric oxygen surrounding**, e.g. for octahedrally coordinated sodium cations (positions SI in zeolite Y, see **Fig. 1**) [1].In the first case, the electric field gradient at the cation sites SII is large, while it is small in the latter case. Therefore, the C_Q value is a characteristic spectroscopic parameter for cations located at different crystallographic cation sites in zeolites. For hydrated zeolites, however, the importance of this spectroscopic parameter is lost, because of an averaging of the local electric field gradients at the different cation sites by mobile water molecules.

An experimental proof of the dominating role of quadrupolar interactions for ²³Na MAS NMR spectroscopy of sodium cations in dehydrated zeolites is given by **Fig. 2**. This Figure shows ²³Na MAS NMR spectra of sodium cations in dehydrated zeolite Na-Y, recorded at Larmor frequencies of $v_0 = 79.3$ to 198.4 MHz, i.e. utilizing magnetic fields of $B_0 = 7.0$ to 17.6 T. These spectra demonstrate that the **center of gravity of the** ²³Na solid-state NMR signals is high-field shifted and the quadrupole patterns become narrow for increasing B_0 values, which is typically caused by the second-order quadrupolar effect. For an explanation of this effect, see e.g. Ref. [12].



Fig. 2

The number and parameters of ²³Na MAS NMR signals contributing to spectra like those in **Fig. 2**, is often investigated by two-dimensional (2D) MQMAS NMR experiments. In **Fig. 3**, the 2D ²³Na ORIACT MQMAS NMR spectrum of a dehydrated zeolite Na-Y with $n_{Si}/n_{AI} = 2.5$ is shown [8]. This spectrum consists of three peaks. See Ref. [11] for details of ORIACT (Off-resonance Rotation-Induced Adiabatic Coherence Transfer) MQMAS experiments.



2D ²³Na ORIACT MAS NMR

Quantitative evaluation of the 2D ²³Na ORIACT MQMAS NMR spectrum in **Fig. 3** delivers the isotropic chemical shifts $\delta_{23Na,iso}$, and the *SOQE* parameters (Second-Order Quadrupolar Effect, see e.g. Ref. [12]) of the signals responsible for the three peaks. These data are helpful for the simulation of one-dimensional ²³Na MAS NMR spectra. **Fig. 4** demonstrates the results of this procedure for the signal separation in the ²³Na MAS NMR spectra of dehydrated faujasite-type zeolites Na-Y and H,Na-Y (both $n_{Si}/n_{AI} = 2.5$). The signal separation shown in **Fig. 3** is an important prerequisite for determining the relative signal intensities and, utilizing the total sodium concentration determined by chemical analysis, the population of sodium cations at the different crystallographically non-equivalent cation sites.

Due to the large number of cation sites in zeolites, such as shown in **Fig. 1** for the faujasite structure, it is a complex topic to attribute the various ²³Na MAS NMR signals to cations at the specific crystallographic sites. A helpful procedure can be https://michael-hunger.de

²³Na MAS NMR studies of cation-exchanged zeolites. In the spectrum of dehydrated zeolite H,Na-Y in **Fig. 4**, **right**, the intensities of the quadrupolar patterns at the right-hand side are much weaker in comparison with those in the spectrum of zeolite Na-Y in **Fig. 4**, **left-hand side**. This finding indicates that the quadrupolar patterns must be due to sodium cations at SII and/or SI' sites, because sodium cations at these sites are preferentially exchanged by hydroxyl protons in faujasite zeolites.



Furthermore, applying the point charge model described in Ref. [13] and using the atomic coordinates, given for the faujasite structure, e.g., in Ref. [14], and oxygen net charges of -0.8 e, the local electric field gradients at the cation sites of zeolite structures and the quadrupole couplings constants of sodium cations located at these sites can be calculated. In **Table 1**, column 2, the results of such

Zeolite/site	Calculation: C _Q /MHz	Experiment: C _Q /MHz	Experiment: η_Q	Experiment: δ/ppm
NaY				
SI	0.1-0.5	ca. 0.1	0	-12
SI'	4.6-5.6	4.8	0.2	-4
SII	3.4-4.2	4.2	0.2	-12
NaX				
SI	0.2	ca. 0.1	0	-6
SI'	3.6-5.2	5.2	0	-19
SII	4.4	4.6	0	-16
SIII	2.2-2.5	ca. 2.4	0.8	-14 to -30

point charge calculations for sodium cations in zeolites Na-Y and Na-X are summarized [3]. In the next step, the comparison of calculated (second column) and experimentally (third column) derived quadrupole coupling constants allows an assignment of ²³Na MAS NMR signals to the various cation sites (first column). Utilizing the above-mentioned combination of experimental and theoretical approaches, the investigation of cation populations of sodium-containing zeolites with a larger number of crystallographically non-equivalent cation sites is possible.

Fig. 5 shows the ²³Na MAS NMR spectra of a dehydrated zeolite Na-X with n_{Si}/n_{AI} = 1.2, recorded at Larmor frequencies of v_0 = 105.8 MHz (bottom) and 158.7 MHz (top) [3]. The high-field range of the spectra consists of two patterns corresponding to C_Q values of ca. 4.6 MHz and 5.2 MHz, which are caused by sodium cations located on SII and SI' sites, respectively. In the low-field range a narrow signal appears due to sodium cations located at SI sites. In contrast to the ²³Na MAS NMR spectra of dehydrated zeolite Na-Y, at least two additional signals of sodium cations located on SIII positions were found in the chemical shift range of δ_{23Na} = -20 ppm to -30 ppm. For a survey on ²³Na solid-state NMR parameters of sodium atoms in powder materials, see Table 8.2 of Ref. [12].



²³Na MAS NMR

Fig. 5

Catalyst preparation: The observation of the quadrupole patterns of sodium cations at crystallographically non-equivalent sites require a dehydration of the powder samples, e.g. via a standard dehydration inside a "sample tube system 1" at the "vacuum line 1", accessible via the link "*In Situ* Solid-State NMR Techniques". The dehydration starts with an evacuation at room temperature for ca. 10 minutes followed by a temperature ramp from room temperature to 393 K within 2 hours. At this temperature, the sample is dehydrated for 2 hours. Subsequently, the temperature is increased up to 723 K within 3 hours and evacuated at this temperature for 12 hours. After this treatment, the sample tube system is closed via the vacuum valve and disconnected from "vacuum line 1", after this line was ventilated with air. The transfer of the dehydrated sample into the MAS NMR rotor is performed without air contact, e.g. in a mini glove box purged with dry nitrogen gas (see Section "mini glove box", accessible via the link "*In Situ* Solid-State NMR Techniques").

²³Na solid-state NMR studies: Due to the quadrupolar interactions of ²³Na nuclei, their single pulse excitation should be performed by less than $\pi/4$ and most suitable by $\pi/8$ pulses. Often, repetition times of t = 1 to 2 s are suitable. For reaching a suitable resolution of the ²³Na MAS NMR spectra, the sample spinning rate has to be as high as possible and at least v_{rot} = 10 kHz. The 2D ²³Na ORIACT MQMAS experiments were performed with a two-pulse $p_1-t_1-p_2$ -sequence and an appropriate phase cycling [8]. The pulse durations p_1 and p_2 were set to 3 to 5 μ s and 25 μ s with radio frequency power levels corresponding to $v_{\rm rf}$ = 150 and 75 kHz, respectively. The value of p_1 was optimized to maximize the relative contribution of the broad signals. The value of p_2 is bound by the RIACT condition, i.e. equal to a quarter of the rotor period. A spectral width of 750 ppm, corresponding to an increment of 6.3 µs for t_1 , was used for the F1 dimension, with a total of 32 increments. The carrier frequency was chosen to be about 4 kHz off-resonance, which is a value suitable for large quadrupolar coupling constants. For each t_1 increment, a number of scans between 20 (typical) and 900 (for very low sodium contents) were used. The repetition time was set to 500 ms [8]. Referencing of the chemical shift was performed to solid NaCl, which has a shift of $\delta_{23Na} = 7.21$ ppm.

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