Characterization of silicon species at catalyst surfaces by ²⁹Si CPMAS NMR

Spectroscopic background: ²⁹Si nuclei have a spin of I = 1/2 and no quadrupole moment. The ²⁹Si isotope has a natural abundance of 4.7 % only and a sensitivity of 3.7 x 10⁻⁴ in comparison with ¹H nuclei (1.0). Therefore, this isotope is at the limit of those candidates for NMR studies of solids, which are possible to investigate without isotopic enrichment. Due to this low NMR sensitivity of ²⁹Si nuclei, the excitation of these nuclei for solid-state NMR studies is performed in some cases by the crosspolarization (CP) experiment (**Fig. 1** [1]). In this pulse experiment, a significant **enhancement of the signal intensities of spins** *S* **with low natural abundance and/or small gyromagnetic ratio** (*S*: ¹³C, ¹⁵N, ²⁹Si etc.) **is possible**.

In the CP experiment, the high population difference $\Delta N_{\rm I}$ of the nuclear energy levels of the ¹H nuclei, which are dipolarly coupled with spins *I*, is utilized for an enhancement of the low population difference $\Delta N_{\rm S}$ of the spins *S* via polarization transfer from spins *I* to spins *S*. An important prerequisite for this $I \Rightarrow S$ polarization transfer is an equalization of the energy levels of the dipolarly coupled spins *I* and *S* in the magnetic field components $B_{1,\rm I}$ and $B_{1,\rm S}$ of long ($\tau = 1$ to 6 ms) radiofrequency pulses, also denoted contact pulses (**Fig. 1**, top). The $\pi/2$ pulse produces spins *I*



Fig. 1

polarization and the contact pulse allows the polarization transfer from spins I to dipolarly coupled spins S, if the following **Hartmann-Hahn condition** is valid [1]:

$$\gamma_1 B_{1,1} = \gamma_S B_{1,S} \tag{1}$$

The parameters in Equ. (1) are the gyromagnetic ratios γ_1 and γ_5 of the spins *I* and *S* and the magnetic field components $B_{1,I}$ and $B_{1,S}$ of the contact pulse for the spins *I* and *S*, respectively. During the detection of the NMR signal of the spins *S*, a weak and long decouple pulse is irradiated at the spins *I* for averaging dipolar *I-S* interactions being responsible for signal broadening. For the polarization transfer during the contact pulse, however, these dipolar *I-S* interactions are necessary. Therefore, the mobility of the *I-S* coupled species under study must be low during the CP experiment. Since the strength of the dipolar *FS* interaction determined the effectivity of the $I \Rightarrow S$ polarization transfer, the signal intensities in ²⁹Si CPMAS NMR spectra don't correlate with the number of ²⁹Si nuclei in a direct manner.

Silicon atoms inside the framework of silicates and aluminosilicates are tetrahedrally oxygen coordinated (Q^4 or Q_4 species), resulting in different silicon environments, such as Si(*n*AI) or Si((4-*n*)OSi,*n*AI) (n = 0 to 4), with typical ²⁹Si chemical shifts (see "method 6" via link "NMR Methods for Heterogeneous Catalysis" and Refs. [2] and [3]). In addition, silicon atoms at the surface of pure or modified silicates and aluminosilicates are bound via oxygen bridges to hydroxyl protons, which can act as adsorptions sites. As an example, Fig. 2 shows the ²⁹Si CPMAS NMR spectra of microporous Silicalite-1 (a) with MFI structure and of mesoporous SBA-15 (b) [4]. The spectrum of the calcined (T = 723 K) Silicalite-1 consists of signals at $\delta_{29Si} = -103$ ppm and -113 ppm due to SiOH groups of Q³ (Si(3OSi,1OH)) silicon species and due to tetrahedrally coordinated framework silicon atoms of Q⁴ (Si(4OSi)) silicon species, respectively. Upon saturation with H₂O, a weak decrease of these signals is observed (**Fig. 2a**), which is explained by a decreased ${}^{1}H \Rightarrow {}^{29}Si$ polarization transfer upon saturation. This is explained by an increase of the ¹H-²⁹Si distance due to an increase of the O-H distance of the corresponding SiOH groups caused by Hbondings to water molecules. Furthermore, a stronger decrease of the signal intensities occurred upon the saturation of Silicalite-1 with CD₃OH (CD₃OH saturated). This finding hints at stronger $CD_3OH/SiOH$ interactions compared with the $H_2O/SiOH$ interactions for Silicalite-1 [4].

The ²⁹Si CPMAS NMR spectrum of calcined (T = 723 K) SBA-15 in Fig. 2b consists of Q^3 (Si(3OSi,1OH)) and Q^2 (Si(2OSi,2OH)) signals at δ_{29Si} = -101 ppm and -92 **ppm**, respectively, and a weak high-field shoulder at $\delta_{29Si} = -110$ ppm due to Q⁴ species [4]. In contrast to Silicalite-1, saturation of SBA-15 with water (H₂O saturated) led to a significant increase of the ²⁹Si CPMAS NMR signals. This effect may be due to an adsorption of water at the SiOH groups of well accessible Q² silicon species in such a manner that the H atoms of these water molecules are able to contribute to the ¹H \Rightarrow ²⁹Si polarization transfer. In this case, the intensity decrease of the Q² signals of SiOH groups involved in H-bondings to water molecules is overcompensated by the contribution of the additional ¹H spins of adsorbed water molecules to the ${}^{1}H \Rightarrow {}^{29}Si$ polarization transfer. This effect cannot occur for Silicalite-1 since this material has no Q² silicon species. The ²⁹Si CPMAS NMR spectrum of SBA-15 saturated with CD₃OH (CD₃OH saturated), on the other hand, shows significantly lower signal intensities compared with the spectrum of calcined SBA-15, which is similar to the finding for methanol-loaded Silicalite-1. Also for SBA-15, therefore, strong CD₃OH/SiOH interactions can be assumed, leading to an increase of the ¹H-²⁹Si distance due to an increase of the O-H distance of the corresponding OH groups caused by H-bondings to methanol molecules [4].



²⁹Si CPMAS NMR

Fig. 2

Depending on the application of siliceous materials, silicon atoms at the surface are bound via oxygen bridges to carbon atoms or silicon atoms in surface-anchored complexes. After reaction with octadecylmethyldichlorosilane, signals of D₁ ($\delta_{29Si} = -4$ ppm), D₂ ($\delta_{29Si} = -7$ ppm), D₃ ($\delta_{29Si} = -10$ ppm) and of various D₄ + D'₄ species ($\delta_{29Si} = -14$ to -22 ppm) are obtained [5-8]. After reaction with octadecyltrichlorosilane, additional signals of T1 ($\delta_{29Si} = -46$ ppm), T'₁ ($\delta_{29Si} = -50$ ppm), T₂ ($\delta_{29Si} = -56$ ppm), T₃ + T'₃ ($\delta_{29Si} = -59$ ppm), and T₄ + T'₄ species ($\delta_{29Si} = -64$ to -70 ppm) occur (see also **Fig. 3** and **Table 1**) [5-8].



T₄

Fig. 3

Structural type	δ _{29Si} /ppm
Q ₂ Q ₃ Q ₄	-91 -101 -110
$D_1 D_2 D_3 D_4 + D'_4$	-4 -7.2 -10 -14 to -21
$ \begin{array}{c} T_1 \\ T_1' \\ T_2 \\ T_3 + T_3' \\ T_4 + T_4' \end{array} $	$-46-50-55.5 (R \ge CH_3)-59 (R \ge CH_3)-64 to -70$

Table 1

For applications as stationary phases in chromatography or as an easy-to-handle catalytic material, MCM-41 silica spheres were modified by grafting with mono- (MF) and trifunctional octadecyl (TF) alkyl chains and endcapping with hexamethyldisilazane [9]. ²⁹Si CPMAS NMR spectroscopy was utilized for determining the surface species, amount of alkyl chain grafting and degree of crosslinking of the attached alkylsilanes. The ²⁹Si CPMAS NMR spectra of the MCM-41 samples grafted with MF and TF are shown in Figs. 4a and 4b [9]. The ²⁹Si chemical shifts of the various species correspond to those in Table 1. The ²⁹Si CPMAS NMR signals at δ_{29Si} = -100 ppm and -109 ppm originate from surface silanol groups (Q³) and Q⁴ species, respectively. The comparison with the ²⁹Si CPMAS NMR spectrum of the calcined MCM-41 sample prior to grafting (spectrum not shown), with a dominant Q^3 signal as well as Q^2 and Q^4 resonances, reveals a complete loss of the Q² signal and a noticeable reduction of the Q³ signal after surface modification with the alkylsilanes and subsequent endcapping. The ²⁹Si CPMAS NMR spectrum of the MCM-41 sample grafted with TF contains signals of T^n groups $(T^n = RSi(OSi)_n(OH)_{3-n})$ with n = 1, 2, 3) between $\delta_{29Si} = -45$ ppm and -70 ppm. These signals hint at the attachment and cross-linking of the C₁₈ chains on the MCM-41 silica surface. Furthermore, intense ²⁹Si CPMAS NMR signals occur at about δ_{29Si} = 14.5 ppm $(R_3 SiOSi = M group)$, which are a superposition of the signals due to the attached monofunctional C_{18} chains and the trimethyl silyl groups from the endcapping reaction [9].



Fig. 4

Catalyst preparation: Most of the ²⁹Si CPMAS NMR studies of silicon atoms at the surface of pure or modified silicates and aluminosilicates are performed with sample materials in the hydrated state.

²⁹Si CPMAS NMR studies:. The ²⁹Si cross-polarization MAS NMR measurements (CPMAS NMR) in Fig. 2 were carried out using a 7.0 mm MAS NMR probe with a spinning rate of $v_{rot} = 3.5$ kHz. These spectra were recorded at the resonance frequency of $v_0 = 79.5$ MHz, with a radiofrequency field of $v_{rf} = 41.67$ kHz, a ¹H–²⁹Si contact pulse of $\tau = 3$ ms, and the repetition time of 4 s. For cross-polarization experiments, the repetition time is limited by the T_1 time of the ¹H nuclei, which is much shorter than for ²⁹Si nuclei. For decreasing the ²⁹Si MAS NMR line width, high-https://michael-hunger.de

power ¹H decoupling was applied In the case of ²⁹Si CPMAS NMR studies, however, the signal intensities do not correlate with the real spin numbers, which hinders the quantitative evaluation of obtained spectra. Chemical shifts are referenced to octakis(trimethylsiloxy)silsesquioxane powder (Q_8M_8 , $\delta_{29Si} = 11.7$ ppm) at spinning rates of $v_{rot} = 2$ to 3 kHz. This reference is easier to handle in MAS NMR rotors in comparison with tetramethylsilane (TMS, $\delta_{29Si} = 0$ ppm) with a boiling point of 299 to 301 K.

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